Biological Sciences Animal Facility Handbook
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Welcome

This handbook has been prepared to provide information and guidelines for anyone currently using or planning to use animals in the Biological Sciences Animal Facility. It may not cover all of your questions, so please feel free to ask any of the people listed in the section below.

The Biological Sciences Animal Facility provides animal care to the University’s Faculty Investigators. The facility is staffed from 8:00 am to 2:00 pm on weekdays and part-time on weekends. The staff is made up of both full-time and part-time staff members, many of which hold American Association for Laboratory Animal Science certifications. The staff is also available to assist researchers and instructors whenever possible, but please keep in mind their busy schedules and provide advanced warning when possible so that the staff can plan to be available to assist you.

Standard animal caging, feed, lighting, temperature and humidity are provided by the facility, exceptions made only with approval from both IACUC and Assistant Director.
Biological Sciences Animal Facility Handbook

University Research Animal Resources

Life Sciences/Veterinary Medicine Unit Staff

Attending Veterinarian/Director: Leanne Alworth
Phone: 542-6084
E-mail: alworth@uga.edu

Animal Resources Manager: Robin Kavanaugh
Phone: 706-542-6083
E-mail: robink@uga.edu

Biological Sciences Facility Staff

Facility Supervisor: Todd H. McDaniel
Phone: 706-542-7443
E-mail: toddmcd@uga.edu

There are also part-time student workers available in the facility.

Emergency numbers are posted next to or above each facility phone. Phones are located:
Facility Main Hallway

Important Phone Numbers
Office: 706-542-7443
Key Cards and Facility Access

The BioSciences Animal facility is protected by a magnetic card reader system. All facility users seeking access to BioSciences animal facility must acquire a proximity card from the UGA Card Office in the Tate Center. Cards should be obtained at least 2 working days before expected use, to enable any unforeseen problems to be cleared before the card is needed.

Once a proximity card is obtained, a legible photocopy should be made of the front and back of the card with the name of the lab’s head investigator written on the copy. The copy should then be given to the facility supervisor, Todd McDaniel. Access to the facility is usually granted in 24 hours. If you do not already have building access, please contact Robin Kavanaugh at (706) 542-6083.

Please keep in mind that children and pets are not permitted in the facility. A member of the staff must accompany all visitors.

Entry and Exit Procedures

Facility Entry
To enter the facility one must pass their proximity card over the card reader. The door will audibly unlock. When entering the animal facility, be sure not to allow others to follow in behind you. This undermines the security of our facility and keeps us from identifying users. Do not let in people you do not recognize, even if they say they need to be let in. If you hear knocking, or someone follows you in, find the Facility Supervisor or a Full-time Technician and alert him or her.

Personal protective equipment (PPE), such as lab coats, scrubs and shoe covers, are available in the main hallway and room 157A. To enter the facility, you must wear shoes with covered toes. If you are entering the facility, but will not enter an animal room or handle equipment, no further PPE is required. To enter an animal room, or handle equipment, you must wear a lab coat, gown, or scrubs. There are PPE signs outside of each animal room, pay close attention to the specific PPE listed for the room you plan on entering.

Entering General Animal Rooms
To enter an animal room you must wear a lab coat, gown or scrubs. This practice is to protect you from carrying allergens and contaminants on your street clothes back to your lab or home, and to protect your mice or rats from the microbes you bring into the facility on your street clothes. Upon entering the animal room you must wear gloves to handle cages or animals. This protects both you and the animals.

Exiting General Animal Rooms
Wash your hands after removing and properly disposing of your gloves in the trash receptacle provided by the door to each room.
Rooms Requiring Special Procedures

Entering a Barrier/Immunocompromised Room
Barrier housed mice are sterile housed and require special procedures to ensure their health. Autoclaved or disposable gowns, gloves, face masks and shoe covers are available inside or immediately outside of the rooms; do not use personal protective equipment (PPE) from other rooms in the facility. Tape on the floor marks the barrier “ante-room” area, where PPE is donned. You can only step into the area beyond the tape with shoe covers. Gowns should be donned before crossing the barrier tape, and shoe covers should be put on one at a time while stepping over the barrier tape one foot at a time. Gloves should be donned after shoe covers have been put on, to keep them from getting contaminated if they touch your shoes.

Exiting a Barrier/Immunocompromised Room
When procedures are finished in the barrier room, you may walk directly over the tape and remove your PPE. Autoclaved gowns can be reused if placed back in their bags. Bags should be labeled with the name of the user, and after one week of use they should be placed in the gown bin to be laundered and re-autoclaved. Gloves, shoe covers and facemask should be thrown away in the trash receptacle provided.

Entering Infected/Quarantine Rooms
Animals with questionable health status and certain infectious diseases are housed in isolation racks in quarantined rooms. Bio-containment is extremely important in these rooms and we require your full cooperation in these procedures. Disposable isolation gowns, facemasks, gloves and shoe covers are provided in the staging area of each room that holds infected or quarantined animals. All of these items must be worn to protect both user and animals. There is no need for step over procedures for shoe covers when entering in these rooms.

Exiting Infected/Quarantine Rooms
Specific exit procedures are required to contain infectious organisms within the quarantined room. Shoe covers should not be allowed to touch the floor beyond the quarantine room threshold, so remove shoe covers as you step over the threshold, placing them into the biohazard trash receptacle provided. Remove the isolation gown and mask and place them into the biohazard trash receptacle provided. Remove your gloves last.

If you have dirty biohazardous cages to remove from the room, they must also be bagged inside the animal room before you remove your PPE. The bag should be sprayed with the appropriate disinfectant. The appropriate disinfectant will be agreed upon by animal resources in consultation with your lab and provided by animal resources. Remember to notify the staff that you have left dirty cages in the room, so they may be autoclaved and properly handled.

Exiting the Animal Facility
Upon finishing your work, you should always exit through the main hallway door. Laundry baskets are provided for dirty lab coats and scrubs on the left as you exit.

Animal Environment

Macro-environment
Light cycles in the animal rooms are controlled by electronic time clocks in each room. The default light cycle is 12:12, 12 hours light and 12 hours of dark. Light cycles can be adjusted
by the Facility Supervisor if an investigator’s research warrants it. If working in BioSciences
after the dark cycle has started, be courteous and make every attempt not to disturb other
investigator’s animals.

Housing
The BioSciences animal facility houses mice and rats. The facility follows all federal
regulations and guidelines for the housing of animals in research. All animals are provided
with 250ml or 500ml Ancare brand water bottles. Bottles are changes weekly and filled with
A-CC municipal tap water. All bottles are checked daily for leakage.

For mice, Ancare brand cages are the standard housing provided by the facility. These cages
are bedded with Bed-o-cob combination bedding and a nestlet. Based on federal regulations,
the maximum number of adults which can be housed in one cage is 5 mice. Please adhere to
this regulation. Standard shoebox caging is provided for rats. Regular water bottles are
provided for drinking water. Smaller rats (up to 300g) may be pair housed, but rats over 300g
must be singly housed if in rat shoebox cages.

Feed is provided in-room, in labeled containers. BioSciences animal facility provides
irradiated Purina Lab Diets, including 5053 – Rodent Diet, 5058 – Mouse Diet (Breeder Diet).
Arrangements for special diets must be made with the Facility Supervisor.

Animal Use

Handling and Animal Use
Any investigator, or researcher may request an animal handling training session from the Life
Sciences Animal Resources Unit if they feel one would benefit their work. Please contact the
Facility Supervisor or Attending Veterinarian regarding handling training. When working with
or around the mice, work quietly to avoid disturbing or distressing the mice. Any excessively
distressing procedures, such as blood collection, euthanasia, any surgeries or animal
manipulations must be done outside of the animal housing room, in the PI Labs or in the
facility procedure/surgery room. These procedures are to protect the animals and to avoid
unnecessary stimulation of the animals.

During the course of your research please keep in mind that mouse cages must only be opened
inside a laminar flow change station or biosafety cabinet. Any and all in-room animal
manipulations must be done under a laminar flow change station or biosafety cabinet. These
procedures are to protect the animals. The biosafety cabinets in the facility are not appropriate
for work with volatile chemicals or anesthetic gases.

Anesthesia
URAR-LS requires researchers who use the animal facility provided anesthesia vaporizers to
provide and use their own induction chamber and nose cone/face mask. URAR does not
provide these items for general use.

This is to protect the health status of the colony rodents by preventing the spread of infection
among colonies in the facility via anesthesia equipment. In order to prevent contamination
from your lab, please do not bring this equipment back and forth from your lab. You will be
allowed to store your items in the animal facility, and your Supervisor will inform you of the

location options for storage. We suggest you keep it in a locked container labeled with your lab name.

The systems are manufactured by VetEquip. Purchase information is listed below. Please ask your Supervisor if you have any questions.


**Breeding**

There are alternate methods for breeding rodents.

Pair breeding consists of 1 male and 1 female in the cage. With this method, only 1 female is reproducing with the male, but due to the post-partum estrus, they can have a litter approximately every 21 days.

Harem/group breeding consists of 1 male with multiple females (2-4 depending on the cage dimensions). We do not allow multiple litters in the same cage, because pups of different ages can out compete each other for milk, or trample each other, and by the time of weaning, the cage is too crowded. Therefore, with harem breeding, females must be separated before they give birth. With this method, more females are simultaneously reproducing with 1 male, but they do not mate at the post-partum estrus, and will not breed again until the litter is weaned and the female is placed back with the male.

**Weaning**

Litters of mice over the age of 21 days are considered adults and should be weaned accordingly, unless specific exceptions are in the project’s AUP to extend the age of weaning. Weanlings should be separated by sex into cages of 5 animals or less, independent of their weight at weaning. If training is needed on weaning procedures, please contact the Facility Supervisor.

**Census Log**

Any permanent removal of animals, animals found dead, newly weaned animals and transfer of animals from another investigator’s colony or vice versa must be recorded on the census sheet to ensure proper billing. Animals and cages are both counted. The personnel responsible for their weaning must add weaned litters to the census sheet. Columns on the left side of the sheet are designated to different categories such as, received/weaned, used by the lab, transferred or found dead. The two large columns are for animal count and cage count. Any entries should be initialed.

**Reporting Sick or Dead Animals**

If a sick or dead animal is found in an animal room, the animal must be reported to the lead tech and/or facility supervisor, or the vet tech. If no one is available, or animals are found on the weekend or a holiday, there are numbers for on-call veterinary assistance listed next to the Facility Supervisor’s office.
Sick animals should be reported to a member of the URAR staff immediately. Sick animals will be given an identifying tag or cage card. For mice, the pink identifying card is the medical record, and all treatments must be noted on the back of the card. For some species, a paper medical record maybe used. The Attending Veterinarian and/or Vet Tech will be contacted, will determine the appropriate veterinary care, and will contact, directly or via the Animal Resources staff, the investigator’s lab with the recommended treatment plan. The plan will be discussed, and with input from the lab, a plan will be decided. It is the responsibility of the lab to follow the treatment plan decided. Changes to the plan may only be made by the Attending Veterinarian or Vet Tech.

Dead animals should be removed from the home cage and placed in a bag labeled with the room number, investigator name, date and location of cage. Once placed in a properly labeled bag, the carcass should be put in the necropsy refrigerator (main hallway) and logged in on the log sheet. The animal should also be recorded under the found dead column of the census sheet on the proper date. The lead tech or facility supervisor should be informed of the death.

**Acquiring Equipment**

Clean cages are provided for your use in Clean Cage Storage (Room 161). You may take clean cages as needed, but please take only the number of cages you need, as cages cannot be used in other rooms once they have entered an animal room. If your animals are housed in a barrier room, or housed in sterile cages, cages are prepared by the animal care staff and provided in the animal rooms. Please notify your facility supervisor if you find you do not have enough prepared equipment available.

**Animal Transportation**

Movement of animals from one animal room to another animal room within this facility or to another animal facility, must be arranged with your facility supervisor before you move the animals. Also, if you plan to take animals out of the facility and return them alive to be housed in the facility again, you must arrange the plan with your facility supervisor before you move the animals.

*Transportation within the Facility*

Animals that are being transported within the facility must be transported in a closed container, preferably a rodent cage. This is to prevent escapes and protect the health of both animals and staff.

*Transportation outside the Facility, within the Building*

Animals that are being transported to labs on the upper floors of BioSciences must be transported in cages with microisolator tops. Use of the home cage is preferred, if possible. All animals are to be sighed out on the log sheet on each animal room’s door.

If live animals are returned to the facility, the container must be sprayed thoroughly with disinfectant before it is placed into the laminar flow change station or biosafety cabinet. The animals should be transferred to a clean cage, and the dirty cage taken directly to the dirty cage wash room.
Transportation out of the Building

Animals that are being transported out of the BioSciences building must be transported in closed opaque containers or containers that have been covered to conceal their contents. Covering the cage with a trash bag is acceptable, along with using larger shopping bags or dark plastic that conceal their contents. Bags should not be closed tightly, and should be removed from the cages as soon as they arrive at their destination, to prevent smothering. For rats or large numbers of mice, BioSciences animal facility staff can provide you with shipping boxes to use as temporary holding during transport to the lab. If live animals are returned to the facility, the container must be sprayed thoroughly with disinfectant before it is placed into the laminar flow change station or biosafety cabinet. The animals should be transferred to a clean cage, and the dirty cage taken directly to the dirty cage wash room.

Animal Health and Veterinary Services

The Attending Veterinarian for University Research Animal Resources- Life Sciences Unit provides veterinary services for the Coverdell Rodent Vivarium.

The animal care staff checks animals visually, on a daily basis. Any injuries or illness noted by the animal care staff will be reported to the Facility Supervisor and/or Attending Veterinarian and/or Vet Tech. A pink Sick animal card and placed on the cage. This card should only be removed when the animal is dead or the illness/injury has been determined to be resolved. Only the Attending Veterinarian or the Vet Tech, or their designee may resolve a case. The Facility Supervisor and/or Attending Veterinarian and/or Vet Tech will examine the animal. Investigators will then be contacted about the animal and the recommended treatment plan.

Concerns about animal welfare, illness or injuries can be taken to your lead technician, Facility Supervisor, or Attending Veterinarian.

Euthanasia Station

BioSciences animal facility has one euthanasia station one located in Necropsy/Procedures (room 157). The station provides gaseous CO2 for humane euthanasia. Standard Operating Procedures for the use of the CO2 stations are posted and must be followed. Federal regulations stipulate that a secondary, physical method is required after CO2 administration to ensure complete euthanasia. Appropriate secondary methods are listed at the stations. The method of euthanasia used must be on the approved research AUP.

If you need instruction on proper euthanasia techniques, please contact the Facility Supervisor.

Non-Animal Investigator Needs

Change Room

The changing room is the facility bathroom, room 160, and has a shower.

If your lab uses gowns, you may use the sets of washers and dryers in the hallway to launder them, but you must get instruction from the Animal Resources staff before using them. We ask that you do not bring lab coats from your personal labs into the facility, as we provide
clean, facility laundered lab coats for your use in the facility. Please let a member of the staff
know if you are unable to locate a clean lab coat.

**Cabinets and Drawers**

Biological Sciences facility does have a small number of locking cabinets and drawers found
in the rodent procedure room and necropsy room 157. If you would like to use one of these
spaces for your lab’s items, please speak with the Facility Supervisor, at least 48 hours in
advance of when you will need the space.

**Drugs**

Investigator drugs should be stored in the investigator’s lab, not the animal facility. If an
investigator feels that s/he must store drugs in the facility, the investigator will need to discuss
the matter with the Facility Supervisor and Attending Veterinarian.